STAINING PROCEDURE

1. For paraffin sections, deparaffinize and hydrate tissue sections through xylenes or other clearing agents and graded alcohol series.

   For frozen sections or cell preparations fix with acetone or an appropriate fixative for the antigen under study, if necessary.

   Wash for 5 minutes in tap water.

2. If antigen unmasking is required, perform this procedure using a Vector® Antigen Unmasking Solution, Citrate-based, pH 6.0 (H-3300) or Tris-based, pH 9.0 (H-3301).

3. If quenching of endogenous peroxidase activity is required, incubate the sections in BLOXALL® Blocking Solution for 10 minutes.

4. Wash in buffer for 5 minutes.

5. Incubate sections for 20 minutes with Normal Horse Serum, 2.5%.

6. Tip off excess serum from sections.

7. Incubate with goat primary antibody diluted in appropriate antibody diluent solution, such as diluted normal horse serum or BSA.

8. Wash in buffer for 5 minutes.

9. Incubate for 30 minutes with ImmPRESS-AP Polymer Reagent.

10. Wash for 2 x 5 minutes in buffer.

11. Incubate in alkaline phosphatase substrate solution (not included) until desired stain intensity develops.

12. Rinse sections in tap water.

13. Counterstain (optional), clear and mount.

Detailed product listings, specifications, protocols and additional information are available on our website: vectorlabs.com