STAINING PROCEDURE

1. For paraffin sections, deparaffinize and hydrate through xylenes or other clearing agents and graded alcohol series.

For frozen sections or cell preparations fix with acetone or an appropriate fixative for the antigen under study, if necessary.

Wash for 5 minutes in tap water.

2. If antigen unmasking is required, perform this procedure using a Vector® Antigen Unmasking Solution, Citrate-based, pH 6.0 (H-3300) or Tris-based pH 9.0 (H-3301).

3. If quenching of endogenous peroxidase activity is required, incubate the slides in BLOXALL® Blocking Solution (SP-6000) for 10 minutes.

4. Wash in buffer for 5 minutes.

5. Incubate for 20 minutes with diluted normal blocking serum.

6. Tip off excess serum from sections.

7. Incubate for 30 minutes with primary antibody diluted in appropriate diluent such as buffer with 2.5% normal serum or R.T.U. Animal-Free Block and Diluent (SP-5035).

8. Wash for 5 minutes in buffer.

9. Incubate for 30 minutes with diluted biotinylated secondary antibody.

10. Wash for 5 minutes in buffer.

11. Incubate for 30 minutes with prepared VECTASTAIN ABC Reagent.

12. Wash for 5 minutes in buffer.

13. Incubate in a peroxidase substrate solution (not included in kit) until desired stain intensity develops.

14. Rinse in tap water.

15. Counterstain (optional), clear and mount.

Detailed product listings, specifications, protocols and additional information is available on our website: vectorlabs.com